

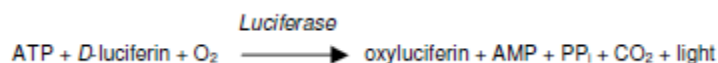
Adenosine Diphosphate (ADP) Assay Kit (Chemiluminescent)

LS-K257-100 (100 Tests) • Store at -20°C



Introduction

This ADP Assay Kit provides a rapid Method to measure ADP levels. The assay involves two steps. In the first step, the working reagent lyses cells to release ATP and ADP. In the presence of luciferase, ATP immediately reacts with the Substrate *D*-luciferin to produce light. The light intensity is a direct measure of intracellular ATP concentration.



In the second step, the ADP is converted to ATP through an enzyme reaction. This newly formed ATP then reacts with the *D*-luciferin as in the first step. The second light intensity measured represents the total ADP and ATP concentration in the sample. This non-radioactive, homogeneous cell-based assay is performed in microplates. The reagent is compatible with all culture media and with all liquid handling systems for high-throughput screening applications in 96-well and 384-well plates.

Key Features

- Safe. Non-radioactive assay.
- Sensitive and accurate. As low as 0.02 μM ADP can be quantified.
- Homogeneous and convenient. "Mix-incubate-measure" type assay. No wash and reagent transfer steps are involved.
- Robust and amenable to HTS: Z' factors of 0.5 and above are routinely observed in 96-well and 384-well plates. Can be readily automated on HTS liquid handling systems for processing thousands of samples per day.

Applications

- ADP determination in cells and other biological samples.

Components

| Component | K257-100 |
|---------------------|-------------------|
| | 100 Tests |
| Assay Buffer | 10 mL |
| Substrate | 120 μL |
| Co-substrate | 120 μL |
| ATP Enzyme | 120 μL |
| ADP Enzyme | 120 μL |
| Standard (3 mM ADP) | 100 μL |

Storage

The kit is shipped on ice. Store all reagents at -20°C. Shelf life: 12 months after receipt.

FOR RESEARCH USE ONLY! Not for use in humans.

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Assay Procedure

Assays can be carried out in a tube or in a 96-well plate. For consistency, it is recommended that the time between the two luminescence measurements be the same for all samples.

Procedure

1. Standard Curve. Prepare 500 μL 30 μM ADP Premix by mixing 5 μL 3 mM Standard and 495 μL distilled water (for cell culture samples dilute ADP in culture media). Dilute standard as follows. Transfer 10 μL standards into wells of a white opaque 96-well plate.

| No | Premix + H ₂ O/media | Vol (μL) | ADP (μM) |
|----|-------------------------------------|-----------------------|-----------------------|
| 1 | 50 μL + 0 μL | 50 | 30 |
| 2 | 40 μL + 10 μL | 50 | 24 |
| 3 | 30 μL + 20 μL | 50 | 18 |
| 4 | 20 μL + 30 μL | 50 | 12 |
| 5 | 15 μL + 35 μL | 50 | 9 |
| 6 | 10 μL + 40 μL | 50 | 6 |
| 7 | 5 μL + 45 μL | 50 | 3 |
| 8 | 0 μL + 50 μL | 50 | 0 |

Samples. Use 10 μL sample per well in separate wells.

For tissue samples, homogenize 20 mg sample in 200 μL of cold phosphate-buffered saline, spin at 12,000 g for 5 min to pellet any debris. Transfer 1-10 μL supernatant to each well and bring the volume to 10 μL with PBS. Test several doses of the sample and choose the readings that are within the standard curve range for ADP calculation.

For suspension cells, transfer 10 μL of the cultured cells (10^3 - 10^4) into a white opaque 96 well plate.

For adherent cells, culture 10^3 - 10^4 cells in white opaque microplate. At the time of assay, remove the culture medium immediately before adding 90 μL ATP Reagent (see below).

2. ATP Reaction. Bring Assay Buffer, Substrate and Co-substrate to room temperature. Thaw enzyme on ice or at 4°C. Fresh Reconstitution is recommended. Store unused reagents including the enzyme at -20°C.

ATP Reagent. For each 96-well, mix 95 μL Assay Buffer with 1 μL Substrate, 1 μL Co-substrate and 1 μL ATP Enzyme. Add 90 μL ATP Reagent to each well and mix by tapping the plate.

After 10 min, read luminescence (RLU A) on a luminometer.

3. ADP Assay. Prepare ADP Reagent: for each 96-well, mix 5 μL dH₂O with 1 μL ADP Enzyme. Immediately following reading RLU A, add 5 μL ADP Reagent to each well and mix by tapping the plate or pipetting up and down. Incubate for 2 minutes at room temperature.

Read luminescence (RLU B) on a luminometer.

Calculation of ADP Concentration

Subtract RLU A from RLU B for standards and samples. Plot the ΔRLU versus ADP concentration for the standards. From the slope of this plot, the Sample ADP concentration can be computed with the following equation:

$$[\text{ADP}]_{\text{sample}} (\mu\text{M}) = \frac{(\text{RLU B})_{\text{sample}} - (\text{RLU A})_{\text{sample}}}{\text{Slope}}$$

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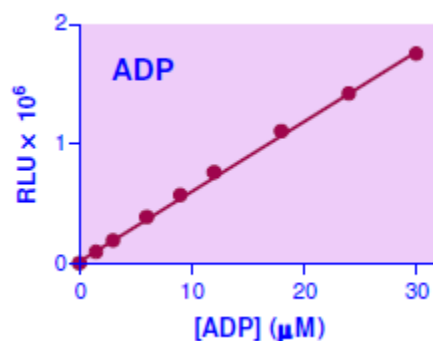
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General Considerations

Signal stability. Since the signal of the reaction decreases by ~1% each minute, for most accurate results, care should be taken that the time between adding the Reconstituted Reagent and luminescence reading is the same for all samples and standards.

Sample Data



ADP Standard Curve in Water

Version: V.08.09.2018

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